



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

Heat shock protein 90 inhibition abrogates TLR4-mediated NF-kB activity and reduces renal ischemia-reperfusion injury

Citation for published version:

O'Neill, S, Humphries, D, Tse, G, Marson, LP, Dhaliwal, K, Hughes, J, Ross, JA, Wigmore, SJ & Harrison, EM 2015, 'Heat shock protein 90 inhibition abrogates TLR4-mediated NF-kB activity and reduces renal ischemia-reperfusion injury', *Scientific Reports*, vol. 5, 12958. <https://doi.org/10.1038/srep12958>

Digital Object Identifier (DOI):

[10.1038/srep12958](https://doi.org/10.1038/srep12958)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Publisher's PDF, also known as Version of record

Published In:

Scientific Reports

Publisher Rights Statement:

This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



SCIENTIFIC REPORTS

OPEN

Heat shock protein 90 inhibition abrogates TLR4-mediated NF- κ B activity and reduces renal ischemia-reperfusion injury

Received: 07 April 2015

Accepted: 06 July 2015

Published: 07 August 2015

Stephen O'Neill¹, Duncan Humphries¹, George Tse¹, Lorna P Marson¹, Kevin Dhaliwal¹, Jeremy Hughes¹, James A Ross², Stephen J Wigmore¹ & Ewen M Harrison¹

Renal ischemia-reperfusion injury (IRI) is a common cause of acute kidney injury. Toll-like receptor 4 (TLR4) mediates sterile inflammation following renal IRI. Heat shock protein 90 (Hsp90) inhibition is a potential strategy to reduce IRI, and AT13387 is a novel Hsp90 inhibitor with low toxicity. This study assessed if pre-treatment with AT13387 could reduce renal IRI and established if the mechanism of protection involved a reduction in inflammatory signalling. Mice were pre-treated with AT13387 prior to renal IRI. 24 h later, renal function was determined by serum creatinine, kidney damage by tubular necrosis score, renal TLR4 expression by PCR and inflammation by cytokine array. *In vitro*, human embryonic kidney cells were co-transfected to express TLR4 and a secreted alkaline phosphatase NF- κ B reporter. Cells were pre-treated with AT13387 and exposed to endotoxin-free hyaluronan to stimulate sterile TLR4-specific NF- κ B inflammatory activation. Following renal IRI, AT13387 significantly reduced serum creatinine, tubular necrosis, TLR4 expression and NF- κ B-dependent chemokines. *In vitro*, AT13387-treatment resulted in breakdown of I κ B kinase, which abolished TLR4-mediated NF- κ B activation by hyaluronan. AT13387 is a new agent with translational potential that reduces renal IRI. The mechanism of protection may involve breakdown of I κ B kinase and repression of TLR4-mediated NF- κ B inflammatory activity.

The rising incidence of acute kidney injury represents a significant global health concern¹. Renal ischemia-reperfusion injury (IRI) is a common cause of acute kidney injury following blood loss, trauma and surgery². Preventative treatments are currently lacking and apart from supportive renal replacement therapy, no other therapies exist for patients who develop acute ischemic kidney injury. The best treatments presently available are avoidance of further kidney damage through careful resuscitation, effective treatment of sepsis and avoidance of nephrotoxic medications³.

It has previously been demonstrated that functional and morphological protection from renal IRI in mice follows pre-treatment with the Heat shock protein (Hsp) 90 inhibitor 17-dimethylamino-ethylamino-17-demethoxygeldanamycin (17-DMAG)⁴. However, the molecular mechanisms of protection from renal IRI offered by Hsp90 inhibitors remain to be fully delineated. Defining these mechanisms are essential for the further development of these agents and translation to patients.

It is now recognised that inflammation is perhaps the most critical pathophysiological process involved in the propagation of renal IRI². Toll-like receptor 4 (TLR4) is a key regulator of the pro-inflammatory transcription factor NF- κ B and plays a dominant role in mediating sterile kidney damage following renal IRI⁵. I κ B kinase (IKK) activation leads to the dissociation of NF- κ B from I κ B and its subsequent

¹MRC Centre for Inflammation Research, University of Edinburgh, Edinburgh, EH16 4SA. ²MRC Centre for Regenerative Medicine, University of Edinburgh, Royal Infirmary of Edinburgh, 49 Little France Crescent, Edinburgh EH16 4SA. Correspondence and requests for materials should be addressed to S.O.N. (email: stephenoneill@doctors.org.uk)

activation⁶. Hsp90 may be needed to stabilise the IKK⁷. Consequently, Hsp90 inhibitors could cause dissociation of the IKK complex, prevention of NF- κ B activation and a reduction in inflammation.

This study investigates the mechanism of protection offered by Hsp90 inhibitors in renal IRI. Specifically, it addresses the hypothesis that Hsp90 inhibition represses TLR4-mediated NF- κ B inflammatory activity through breakdown of IKK. AT13387 (Astex Pharmaceuticals) is a new small molecule Hsp90 inhibitor with a low toxicity profile and better translational potential than 17-DMAG. Therefore this study also aims to determine the efficacy of AT13387 in reducing renal IRI⁸.

Methods

***In vivo* experiments.** *Animals.* Male FVB/n mice aged 6–8 weeks and weighing 20–25 g, were used from in-house colonies. Mice were allowed free access to standard chow and water. The mice were kept in a 12:12-h light-dark cycle. All work involving animals was conducted in accordance with the provisions of the UK Animals (Scientific Procedures) Act 1986. The United Kingdom Home Office approved all experimental protocols. Procedures were performed under Home Office project license 60/3737 and personal license 60/3999.

Drugs. The Hsp90 inhibitor AT13387 was kindly provided by Astex Pharmaceuticals⁹. For *in vivo* experiments AT13387 (80 mg/kg free-base equivalent) was dissolved in (2-Hydroxypropyl)- β -cyclodextrin (2H β C) (Sigma-Aldridge, Dorset, UK), which was prepared in distilled water at 17.5% weight per volume. Mice were treated by intra-peritoneal injection 24 h prior to surgery. A 2H β C vehicle-treated group of mice served as the controls.

Renal IRI model. An established model of warm IRI of the left kidney together with a right nephrectomy was used¹⁰. The model was designed to inflict a moderate to severe acute tubular injury with zero animal mortality. Mice were anaesthetised with an intra-peritoneal injection of Ketamine (75 mg/kg, Vetalar; Pfizer, Kent, UK) and Medetomidine (1 mg/kg, Domitor, Pfizer). A heated mat maintained body temperature with homeostatic control via a rectal temperature probe (Harvard Apparatus, Boston, MA). A mid-line laparotomy was performed for access to both kidneys through a single incision. The right ureter and renal pedicle were ligated then divided. The right kidney was then removed as a control and placed in Methacarn (70% methanol, 20% chloroform, 10% acetic acid). The pedicle of the left kidney was dissected and occluded using an atraumatic vascular clamp (Micro Serrefine, Fine Science Tools, Linton, UK) for 20 minutes. Following removal of the left pedicle clamp, reperfusion was confirmed visually before closure of the abdomen. The anaesthetic was reversed with Atipamezole (1 mg/kg, Antisedan, Pfizer), and a subcutaneous injection of 0.9% saline (25 ml/kg) and Buprenorphine (0.1 mg/kg) was administered. Animals were recovered in an incubator at 29°C until tissue collection 24 h later.

Tissue collection. Under terminal general anaesthesia, blood was recovered by intra-cardiac puncture, and the serum was stored at –20°C. Kidneys were placed immediately into Methacarn, RNAlater (Life Technologies, Paisley, UK) or were snap frozen.

Scoring of morphological kidney injury. Hematoxylin and eosin-stained sections were evaluated for tubulointerstitial injury. A series of non-overlapping fields (x200) were examined by an observer blinded to the sample number. A scoring system based on the proportion of tubules with necrotic/detached cells was used (tubular necrosis score: 0, none; 1, <30%; 2, 30–70%; 3, >70%)⁴.

Serum creatinine determination. Plasma samples were prepared from whole blood. Creatinine was determined using the creatininase/creatinase specific enzymatic method, utilising a commercial kit (Alpha Laboratories Ltd, Eastleigh, UK) adapted for use on a Cobas Fara centrifugal analyser (Roche Diagnostics Ltd, Welwyn Garden City, UK)¹¹.

Cytokine array panel. Kidney protein lysates (200 μ g per sample) were mixed with biotinylated detection antibodies. The sample/antibody mixture was then incubated with a mouse cytokine array panel (R&D Systems, Abingdon, UK). The panel consisted of a nitrocellulose membrane with capture antibodies for 40 murine cytokines spotted on its surface in a duplicate manner. Cognate immobilized capture antibodies on the membrane bound any cytokine/detection antibody complexes present. Following a wash to remove unbound material, Streptavidin-HRP and chemiluminescent detection reagents were added sequentially followed by development using autoradiography. Light produced at each spot was used to determine the amount of cytokine bound. Mean gray values were quantified using Image J (National Institutes of Health, USA)¹².

Real-time reverse transcriptase-PCR. For the detection of renal TLR4 expression, RNA was extracted from kidneys and purified by TRIzol (Invitrogen, Life Technologies, Paisley, UK). RNA concentration and quality was determined using a Nanodrop spectrophotometer and Agilent Bioanalyzer. Across all samples the mean 260/280 ratio was 2.06 \pm 0.01, and mean RNA integrity number was 7.9 \pm 0.6. cDNA was synthesized using a RT² First Strand Kit (Qiagen, Manchester, UK). Real-time PCR was then performed using a RT² qPCR primer assay for mouse TLR4 (Qiagen, Manchester, UK). Glyceraldehyde

3-phosphate dehydrogenase (GAPDH) was used as the reference gene, and was also quantified using a RT² qPCR primer assay (Qiagen, Manchester UK).

Power calculation. For animal experiments in the renal IRI model, power calculations were performed and the number of mice was kept to a minimum required to answer the research question. Creatinine was selected as the primary outcome measure since it is objectively measured and less affected by hydration status than blood urea nitrogen. Although creatinine levels are affected by muscle mass and thus differ according to mouse gender and age, all experiments were performed on male mice of the same age. Previous data was used to the estimate effect size for the potential reduction in serum creatinine. Based on the reduction in creatinine in animals treated with Hsp90 inhibitors that underwent contralateral nephrectomy and 30 minutes renal IRI, an effect size of 1.70 was calculated⁴. With α set at 0.05 and β set at 0.1 this translated to a required sample size of 9 animals per treatment group¹³.

In vitro experiments. *Cell culture.* Cell lines included human embryonic kidney cells (HEK293) (European Collection of Cell Cultures, Porton Down, UK), stably transfected HEK293 expressing TLR4 (HEK293-TLR4) and stably co-transfected HEK293 expressing both TLR4 and a secreted alkaline phosphatase (SEAP) reporter under the transcriptional control of NF- κ B (HEK293-TLR4-NF- κ B) (Imgenex, San Diego, USA). Cells were maintained in Dulbecco's modified eagle's medium (Gibco, Paisley, UK) supplemented with 10% foetal bovine serum, penicillin (50 U/ml), streptomycin (50 μ g/ml) and non-essential amino acids (5%). The selection agent for HEK293-TLR4 cells was 10 μ g/ml blasticidin (Invivogen, San Diego, CA) and for HEK293-TLR4-NF- κ B cells were 10 μ g/ml blasticidin, 2 μ g/ml puromycin, 200 μ g/ml zeocin and 500 μ g/ml G418/geneticin (Invitrogen, Carlsbad, CA).

Drugs and reagents. 17-DMAG was purchased from InvivoGen (San Diego, CA). For *in vitro* experiments, stock solutions were formed for 17-DMAG and AT13387 in Dimethyl sulfoxide (DMSO) (Sigma-Aldridge, Dorset, UK) and further diluted prior to use. DMSO vehicle treated cells served as the control. Pre-treatment with Hsp90 inhibitors was performed in a time window 6 or 12 h prior to ligand stimulation.

Reagents. TLR4 grade endotoxin-free hyaluronan was purchased from Enzo Life Sciences (Exeter, UK) and polymyxin B from SERVA (Heidelberg, Germany).

Transfection. Transient NF- κ B transfections were performed using Eugene HP (Roche, Lewes, East Sussex, UK) in the experiment assessing NF- κ B activation in HEK293 cells and HEK293-TLR4 cells, as these cells did not have a stably transfected NF- κ B reporter. Transfection efficiency of the NF- κ B reporter construct (GL4.32 [luc2P/NF- κ B-RE/Hygro]) was controlled by co-transfecting with a control vector (pGL4.73 control vector [hRluc/SV40]) (Promega, Southampton, UK).

Western blot. Western blotting was performed as previously described⁴. Briefly, whole cells extracts were produced and protein concentration determined (BioRad, Hemel Hempstead, UK). Proteins were separated by SDS-PAGE (10% Tris/HCl gels) and transferred to nitrocellulose (BioRad, Hemel Hempstead, UK). Nitrocellulose membranes were soaked in blocking buffer then primary antibody. Primary antibodies were from the IKK isoform antibody kit (Cell Signaling Technology, Boston, MA). After washing, the membranes were exposed to a horseradish peroxidase-conjugated secondary antibody. Enhanced chemiluminescence reagent (Amersham, Chalfont St Giles, UK) was applied followed by development with autoradiography. Equality of loading was confirmed by probing for β -actin (BD Biosciences, San Diego, CA). Autoradiography films were uploaded to a Gel Doc (Bio-Rad, Hemel, UK) using Quantity One software before being cropped. The original autoradiography is available in the Supplementary Material.

Flow cytometry. Cells were trypsinised, collected and incubated at room temperature with 2% rabbit serum. Further incubation was then performed with either PBS/0.5% Bovine Serum Albumin (BSA)/0.1% azide (unstained control), mouse IgG1 isotype control antibody (Serotec, Oxford, UK) or purified mouse anti-human TLR4 antibody (BD Pharmingen, San Diego, CA) for test samples (0.5 mg/ml at 1:50 dilution or 0.5 μ g per test sample). Cells were washed then incubated with either PBS/0.5% BSA/0.1% azide for unstained controls or fluorescein isothiocyanate (FITC)-conjugated goat anti-mouse IgG (Silenus Laboratories, Victoria, Australia) for IgG1 control samples and TLR4-test samples. Cells were washed again and results acquired on a BD Accuri C6 flow cytometer (BD Biosciences, San Jose, USA).

Luciferase assay. Luciferase assays were used to investigate NF- κ B activity following transient NF- κ B transfections in HEK293 and HEK293-TLR4 cells. At 24 h after transfection and following experimentation, cells were processed for luciferase activity using the Dual-Glo luciferase assay system and Turner Biosystems Modulus Microplate analyser (Promega, Southampton, UK). Firefly luciferase luminescence activity was normalised to that of the co-expressed renilla luciferase to determine the fold induction and indicate NF- κ B activity. Maximal signal intensity was achieved 3 h following ligand stimulation with hyaluronan.

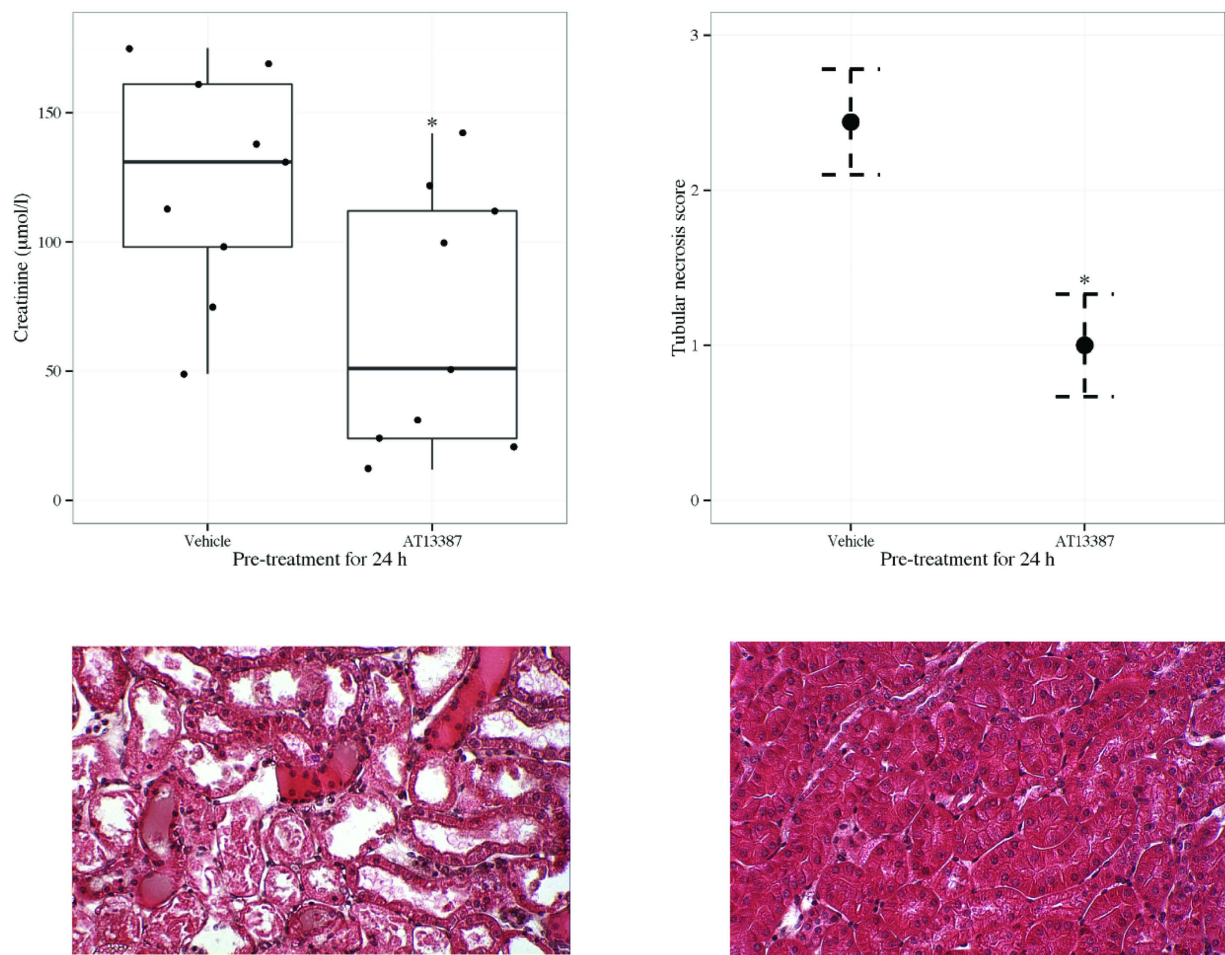


Figure 1. (a) Serum creatinine following AT13387 or vehicle pre-treatment and 24h following renal IRI in FVB/n mice (upper left panel). FVB/n mice were pre-treated with AT13387 or 2H β C vehicle (n = 9 per group). 24h later, mice were anaesthetised and underwent right nephrectomy and 20 min of left renal pedicle clamping. Following 24h of recovery, blood was obtained by intra-cardiac puncture and serum creatinine was determined. Results are presented in a standard boxplot with individual results jittered. * $p < 0.05$ vs. 2H β C vehicle, t-test. (b) Tubular necrosis score following AT13387 or vehicle pre-treatment and 24h following renal IRI in FVB/n mice (upper right panel). The left kidney was harvested and placed in methacarn. Sections were later prepared and stained with haematoxylin and eosin. A blinded observer then determined the tubular necrosis score. Results are presented as mean and standard error of the mean. * $p < 0.05$ vs. 2H β C vehicle, Mann-Whitney U test. (c) Morphological kidney injury following pre-treatment with vehicle and 24h following renal IRI in FVB/n mice (lower left panel). A representative section of kidney cortex at x 200 magnification is shown. (d) Morphological kidney injury following pre-treatment with AT13387 and 24h following renal IRI in FVB/n mice (lower right panel). A representative section of kidney cortex at x 200 magnification is shown.

SEAP assay. SEAP assays were used to investigate NF- κ B activity in HEK293-TLR4-NF- κ B cells as these cells were stably transfected with a SEAP reporter under the transcriptional control of NF- κ B. SEAP assays were performed using the SEAPorter assay kit (Imgenex, San Diego, USA). The concentration of SEAP secreted into cell culture supernatant was calculated from a SEAP standard curve and was used to indicate NF- κ B activation. The assay was performed 24h after the addition of ligands as per the manufacturer's instructions in order to allow adequate time for SEAP to be secreted from cells into the culture medium.

Statistical analysis. Data are presented as mean and standard error of the mean or in standard boxplots with individual results jittered. Statistical comparisons for parametric continuous data were made using student's t-test, one-way analysis of variance (ANOVA) and two-way ANOVA without interaction (using the Tukey's HSD post hoc correction for multiple comparisons). Statistical comparisons for

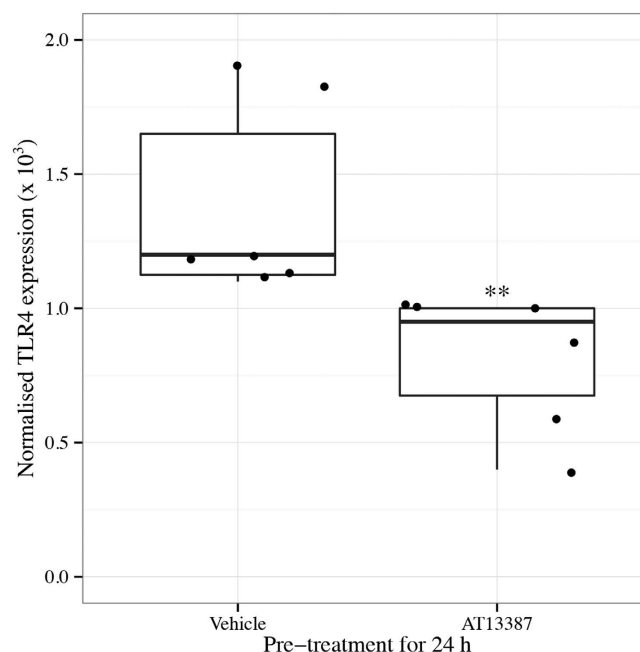


Figure 2. Renal TLR4 expression following AT13387 or vehicle pre-treatment and 24h following renal IRI in FVB/n mice. FVB/n mice were pre-treated with AT13387 or 2H β C vehicle (n = 6 per group) and underwent renal IRI as per Fig. 1. The left kidney was harvested and stored in RNAlater for 24h before being frozen. RNA was later extracted and converted to cDNA. PCR was then performed to determine TLR4 expression. TLR4 expression was normalised to GAPDH expression. Results are presented in a standard boxplot with individual results jittered. **p < 0.01 vs. 2H β C vehicle, Mann-Whitney U test.

non-parametric data were made using the Mann-Whitney U test. All comparisons were performed in R v3.0.1 (R Foundation for Statistical Computing).

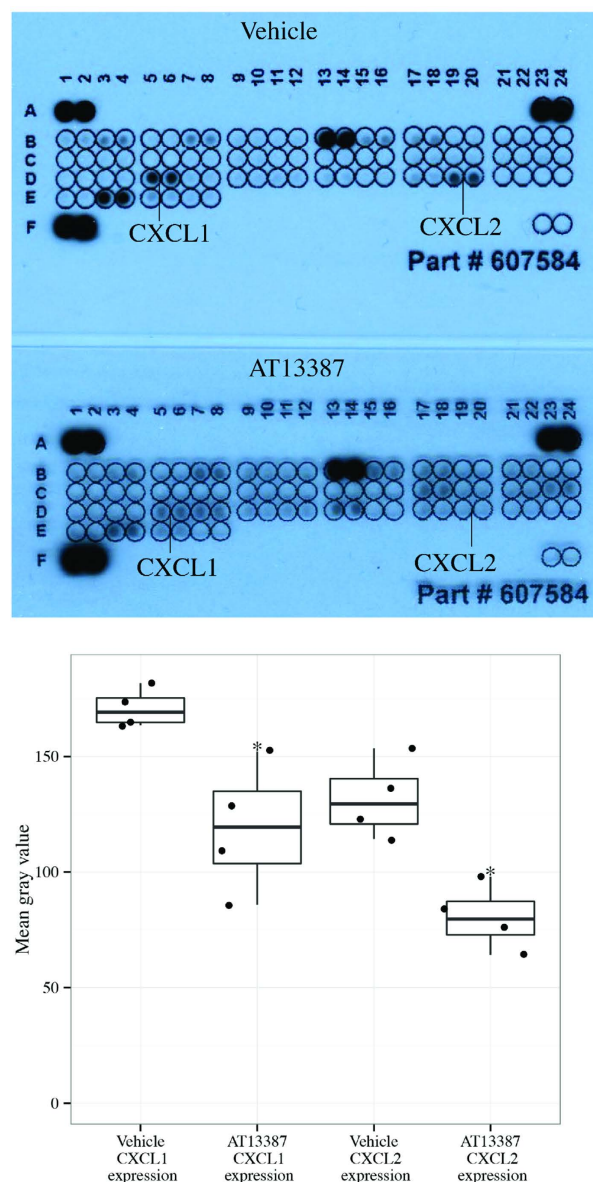
Results

AT13387 reduces renal IRI and inflammatory signalling in the kidney. *AT13387 pre-treatment reduces functional and morphological kidney injury from renal IRI.* On serum analysis, in comparison to vehicle, AT13387 pre-treatment in FVB/n mice significantly reduced creatinine 24h following renal IRI (AT13387 vs. 2H β C vehicle, p < 0.05, t-test) (Fig. 1a). AT13387 pre-treatment in FVB/n mice also reduced tubular necrosis score (AT13387 vs. 2H β C vehicle, p < 0.05, Mann-Whitney U test) (Fig. 1b) on histological assessment 24h following renal IRI (Fig. 1c,d).

AT13387 pre-treatment reduces the expression of TLR4 and inflammatory chemokines in the kidney following renal IRI. On PCR analysis, in comparison to vehicle, AT13387 pre-treatment in FVB/n mice significantly reduced renal TLR4 expression 24h following renal IRI (AT13387 vs. Vehicle, p < 0.01, Mann-Whitney U test) (Fig. 2). On cytokine array panel, in comparison to vehicle, AT13387 pre-treatment in FVB/n mice also reduced renal expression of chemokine (C-X-C motif) ligand 1 (CXCL1) and chemokine (C-X-C motif) ligand 2 (CXCL2) 24h following renal IRI (AT13387 vs. 2H β C vehicle, p < 0.05, t-test) (Fig. 3). There were no other significant differences in cytokine or chemokine expression.

Development of an *in vitro* model of sterile inflammatory signalling. *Selection of a cell line.* The absence of TLR4 expression by HEK293 cells was confirmed by flow cytometry (Fig. 4a). Successful TLR4 transfection was validated in HEK293-TLR4 cells by flow cytometry (Fig. 4b).

Selection of a TLR4-specific and sterile inflammatory ligand. TLR4 was selectively expressed using TLR4-transfected cells (HEK293-TLR4). After ligand stimulation NF- κ B activity was assessed by luciferase assay in comparison to TLR4-null cells (HEK293). Following hyaluronan stimulation there was increased NF- κ B activity in HEK293-TLR4 cells, but an absence of NF- κ B up-regulation in HEK293 cells (HEK293-TLR4 vs. HEK293, p < 0.001, t-test) (Fig. 5a). This indicates that hyaluronan is a TLR4-specific ligand in HEK293-TLR4 cells. The level of TLR4-mediated NF- κ B activation assessed by SEAP assay following hyaluronan stimulation was not altered by the presence of polymyxin B, a drug that clears endotoxin contamination (Fig. 5b).



Coordinate	Target/Control	Alternate Nomenclature
A1, A2	Reference Spot	—
A23, A24	Reference Spot	—
B1, B2	BLC	CXCL13/BCA-1
B3, B4	C5/CSa	Complement Component 5a
B5, B6	G-CSF	—
B7, B8	GM-CSF	—
B9, B10	I-309	CCL1/TCA-3
B11, B12	Eotaxin	CCL11
B13, B14	slCAM-1	CD54
B15, B16	IFN- γ	—
B17, B18	IL-1 α	IL-1F1
B19, B20	IL-1 β	IL-1F2
B21, B22	IL-1ra	IL-1F3
B23, B24	IL-2	—
C1, C2	IL-3	—
C3, C4	IL-4	—
C5, C6	IL-5	—
C7, C8	IL-6	—
C9, C10	IL-7	—
C11, C12	IL-10	—
C13, C14	IL-13	—
C15, C16	IL-12 p70	—
C17, C18	IL-16	—
C19, C20	IL-17	—
C21, C22	IL-23	—
C23, C24	IL-27	—
D1, D2	IP-10	CXCL10/CRG-2
D3, D4	I-TAC	CXCL11
D5, D6	KC	CXCL1
D7, D8	M-CSF	—
D9, D10	JE	CCL2/MCP-1
D11, D12	MCP-5	CCL12
D13, D14	MIG	CXCL9
D15, D16	MIP-1 α	CCL3
D17, D18	MIP-1 β	CCL4
D19, D20	MIP-2	CXCL2
D21, D22	RANTES	CCL5
D23, D24	SDF-1	CXCL12
E1, E2	TARC	CCL17
E3, E4	TIMP-1	—
E5, E6	TNF- α	—
E7, E8	TREM-1	—
F1, F2	Reference Spot	—
F23, F24	PBS (Negative Control)	Control (-)

Figure 3. Renal cytokine expression following AT13387 or vehicle pre-treatment and 24h following renal IRI in FVB/n mice. FVB/n mice were pre-treated with AT13387 or 2H β C vehicle (n = 4 per group) and underwent renal IRI as per Fig. 1. Following 24h of recovery, the left kidney was harvested and snap frozen. Protein lysates were later prepared and an array panel was used to determine cytokine expression. The grid describes the cytokines assessed (right panel). A representative array is shown (upper left panel). Mean gray values were quantified using Image J and were used to reflect CXCL1 and CXCL2 expression. Results are presented in a standard boxplot with individual results jittered (lower left panel). *p < 0.05 vs. 2H β C vehicle, t-test.

Hsp90 inhibition represses sterile inflammatory signalling. *Hsp90 inhibition abrogates TLR4-mediated NF- κ B inflammatory activation.* In HEK293-TLR4-NF- κ B cells, pre-treatment with AT13387 and 17-DMAG significantly reduced hyaluronan-mediated NF- κ B activity assessed by SEAP assay (AT13387 1000 nM vs. DMSO vehicle, p < 0.001, 17-DMAG 1000 nM vs. DMSO vehicle, p < 0.001, ANOVA). In fact, pre-treatment with AT13387 reduced NF- κ B activity assessed by SEAP assay following hyaluronan stimulation to a level equivalent of cells in basal conditions in normal culture medium. In addition, AT13387 was significantly more effective at reducing hyaluronan-mediated NF- κ B activity assessed by SEAP assay than 17-DMAG (AT13387 1000 nM vs. 17-DMAG 1000 nM, p < 0.01, ANOVA) (Fig. 6).

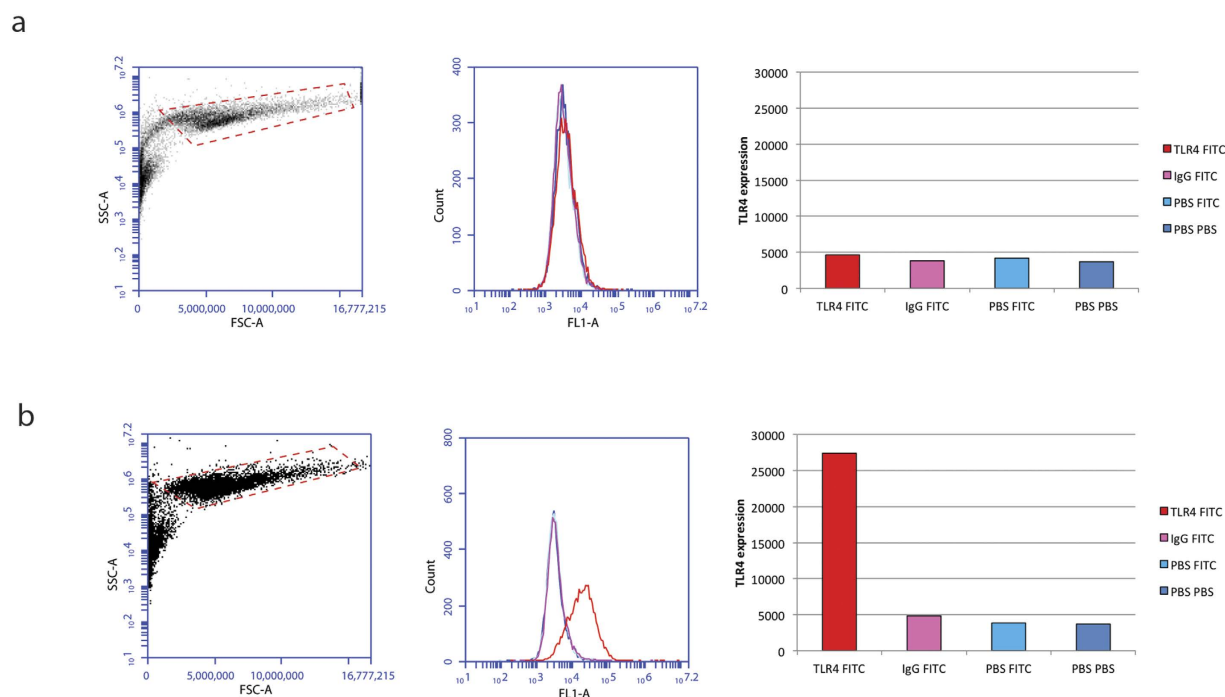


Figure 4. (a) TLR4 expression by HEK293 cells on flow cytometry (upper panel). Cells were trypsinised and collected at 50,000 cells per sample. After processing samples, results were acquired on a BD Accuri C6 flow cytometer. Cells were gated on the basis of forward (FSC-A) and side scatter (SSC-A). TLR4 expression (FLA-1) in HEK293 cells is represented by the red histograms, IgG-FITC negative control sample by the pink histograms, PBS-FITC negative control sample by the light blue histograms and PBS-PBS negative control sample by the dark blue histograms. (b) TLR4 expression by HEK293-TLR4 cells on flow cytometry (lower panel). Cells were analysed and results are reported as per Fig.4a.

Pre-treatment with AT13387 and 17-DMAG leads to breakdown of the IKK complex. Hsp90 inhibition, using pre-treatment with AT13387 and 17-DMAG led to breakdown of the IKK complex on Western blot following hyaluronan stimulation in HEK293-TLR4-NF- κ B cells (Fig. 7 and original autoradiography in Supplementary Fig. 1).

Discussion

In this study, AT13387 was tested in an established renal IRI model to assess efficacy of this agent in reducing renal IRI¹⁰. The experiments performed confirmed that AT13387 pre-treatment leads to both significant functional and morphological protection from renal IRI. This novel finding highlights AT13387 as a potentially exciting new therapy for reducing renal IRI.

Renal IRI predictably occurs in a range of clinical settings including surgery and vascular interventions. It can lead to an acute ischemic kidney injury, potentially life-threatening renal failure, as well as remote organ injury and multi-organ failure. Preventative treatment with AT13387 could be administered prior to anticipated ischemic insults, in individuals at high-risk of acute ischemic kidney injury (e.g. patients with pre-existing kidney disease, diabetes, or previous toxic drug and radiological contrast exposure).

Drugs designed to reduce renal IRI have an absolute requirement for low toxicity. Therefore, one of the main challenges going forward is to develop low toxicity drugs that can be safely utilised in humans. AT13387 is a novel small molecule Hsp90 inhibitor with a low toxicity profile in phase II human studies in oncology and therefore better translational potential than 17-DMAG in this context⁸. Furthermore, since there is patient safety data already available for AT13387, it may be possible to more rapidly translate this therapy into a clinical trial. However, prior to this an increased understanding of the mechanisms of protection offered by Hsp90 inhibition is required.

CXCL1 and CXCL2 are NF- κ B target genes, and the expression of these pro-inflammatory chemokines is NF- κ B dependent^{14,15}. It has previously been identified that following renal IRI, there is increased expression of CXCL1 and CXCL2. Treatment with neutralizing antibodies to both CXCL1 and CXCL2 significantly improves kidney function within 48 hours of renal IRI¹⁶. The expression of CXCL1 and CXCL2 in the kidney was reduced following renal IRI in mice pre-treated with AT13387. There was also a significant reduction in renal TLR4 expression. This suggests an anti-inflammatory effect of AT13387. However, at just 24 hours following renal IRI, it is likely that protective effect of AT13387 may have

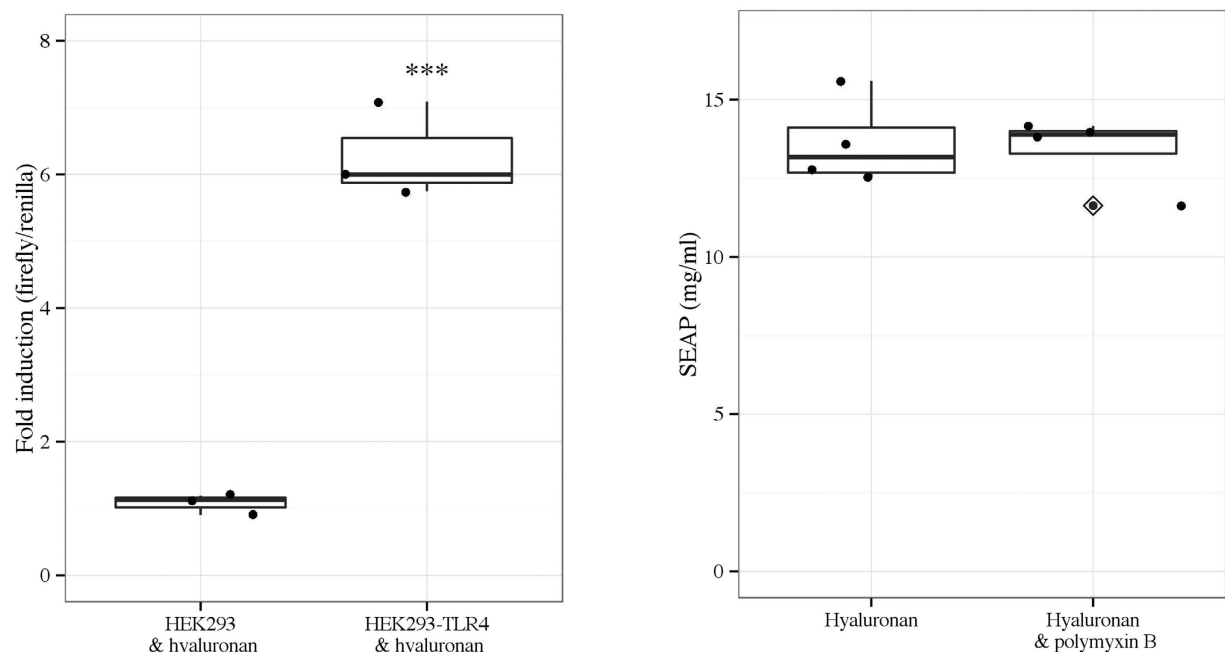


Figure 5. (a) Luciferase assay to determine NF- κ B activity following NF- κ B transfection and hyaluronan stimulation in HEK293 and HEK293-TLR4 cells (left panel). HEK293 and HEK293-TLR4 cells were divided at a cell density of 100,000 cells in 1 ml of growth medium per well of a 12-well plate. After 24 h, cells were transfected with 0.5 μ g pGL4.32 [luc2P/NF- κ B-RE/Hygro] plasmid DNA (experimental vector) using a ratio of DNA to Fugene HP of 3:1. A constitutively active control vector (pGL4.73 [hRluc/SV40]) was added at a ratio of experimental to control vector of 100:1. 24 h later, cells were treated with hyaluronan at a dose of 25 μ g/ml. After 3 h incubation a luciferase assay was performed to determine firefly activity and renilla activity. Fold induction was calculated from the ratio of firefly to renilla and used to reflect NF- κ B activity. Results are presented as 3 independent experiments in a standard boxplot with individual results jittered. The mean fold induction in HEK293 cells was 1.07 \pm 0.09 (standard error of the mean), i.e. there was no increase in NF- κ B activity following hyaluronan stimulation. *** p < 0.001 vs. HEK293 & hyaluronan, t-test. (b) SEAP assay to determine NF- κ B activity following hyaluronan stimulation with and without the presence of polymyxin B in HEK293-TLR4-NF- κ B cells (right panel). HEK293-TLR4-NF- κ B cells were divided at a cell density of 10,000 cells in 100 μ l of growth medium per well of a 96-well plate. 24 h later, the medium was changed to medium containing 25 μ g/ml of hyaluronan either with or without 50 μ g/ml of polymyxin B. After 24 h incubation a SEAP assay was performed to determine NF- κ B activity. Results are presented from 4 independent experiments in a standard boxplot with individual results jittered. Outlier data are highlighted by a dot with a diamond in the midline.

involved programmed cell death rather than inflammation¹⁷. Therefore further mechanistic work was undertaken, and a carefully designed *in vitro* model was developed to simulate the sterile inflammatory environment of renal IRI.

HEK293 cells were selected specifically for these experiments since they transfect efficiently and have previously been reported to lack TLR4 expression^{18,19}. This is in contrast to human renal tubular epithelial cells, which express basal levels of TLR4²⁰.

HEK293 and HEK293-TLR4 cells were therefore considered ideal for identifying a suitable TLR4-specific ligand to explore *in vitro* the hypothesis that Hsp90 inhibition leads to repression of TLR4-mediated NF- κ B inflammatory activation.

Hyaluronan was selected to stimulate the cells, as it is a damage associated molecular pattern molecule and a proposed endogenous TLR4 ligand that is released in increasing amounts during the sterile insult that comprises renal IRI⁵. Hyaluronan was characterised as a sterile TLR4-specific ligand in HEK293-TLR4 cells, and was therefore used to model sterile TLR4-specific NF- κ B activation in further experiments. It was found that Hsp90 inhibition with AT13387 and 17-DMAG resulted in breakdown of IKK α , IKK β and NEMO, which are the three subunits of the IKK complex²¹. This reduced highly specific TLR4-mediated NF- κ B inflammatory activation by hyaluronan to the level of untreated cells left in basal conditions in normal culture medium.

Hsp90 inhibition has previously been used to experimentally treat TLR4-mediated autoimmune diseases^{22,23} and reduce tumour necrosis factor- α -mediated NF- κ B activation^{24–28}. However, this is the

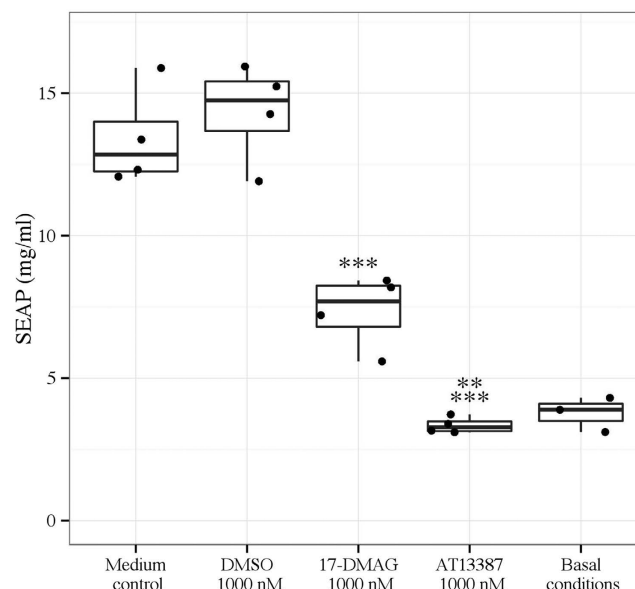


Figure 6. NF- κ B activity determined by SEAP assay following pre-treatment with AT13387 or 17-DMAG and hyaluronan stimulation in HEK293-TLR4-NF- κ B cells. Cells were divided as per Fig. 5b. 24 h later, cells were pre-treated with AT13387, 17-DMAG, DMSO vehicle or medium control for 6 h. With the exception of cells left in basal conditions, hyaluronan was added at a dose of 25 μ g/ml for 24 h and a SEAP assay was performed to determine NF- κ B activity. Results are presented from 3–4 independent experiments in a standard boxplot with individual results jittered. ** $p < 0.01$ vs. 17-DMAG and *** $p < 0.001$ vs. DMSO vehicle, ANOVA.

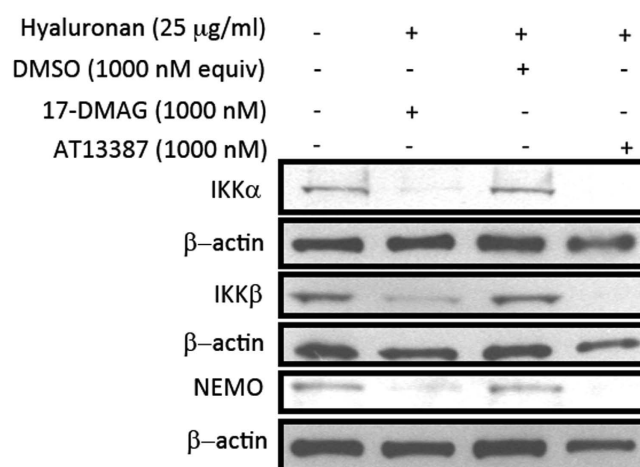


Figure 7. IKK α , IKK β and NEMO levels on Western blotting following AT13387 or 17-DMAG pre-treatment and hyaluronan stimulation in HEK293-TLR4-NF- κ B cells. HEK293-TLR4-NF- κ B cells were divided at a cell density of 500,000 cells in 2 ml of growth medium per well of a 6-well plate. 24 h later, cells were then pre-treated with AT13387, 17-DMAG, DMSO vehicle or medium control for 12 h. With the exception of cells pre-treated with medium control, hyaluronan was added for 4 h. Whole-cell lysates were then prepared and analysed by Western blotting using antibodies to IKK α , IKK β and NEMO, with β -actin being indicated as a loading control. Three different β -actin levels are presented because three separate Western blots were performed on the same samples to avoid antibody interactions. Cropped blots are displayed from each Western blot performed and show protein expression levels of IKK α , IKK β and NEMO in the same samples. The gels were run under the same experimental conditions and original autoradiography is included in Supplementary Fig. 1.

first study to identify that TLR4 signalling to NF- κ B can be targeted by Hsp90 inhibitors. This finding therefore increases our current understanding of the potential role of Hsp90 in renal IRI, as well as inflammatory processes more generally.

In experimental renal IRI, transgenic mice that are missing TLR4^{5,29–31} or that have been treated with TLR4-blocking agents demonstrate a markedly protected phenotype³². Furthermore, de-activation of NF- κ B with small interfering RNAs or decoy oligodeoxynucleotides targeting either IKK β or NF- κ B transcription factors, protects the kidney from IRI^{33–35}.

The potential of Hsp90 inhibitors to interrupt TLR4-mediated NF- κ B signalling is thus highly attractive from a translational perspective since it offers a pharmacological strategy to temporarily dampen NF- κ B inflammatory activity. This is in contrast to ablative strategies targeting IKK that reduce NF- κ B-mediated inflammation but also prevent subsequent NF- κ B-mediated protection from apoptosis³⁶.

Although numerous pathways can lead to NF- κ B activation, IKK is a common point of convergence and activation of NF- κ B occurs nearly universally via IKK-mediated degradation of I κ B⁶. Indeed, it has previously been shown in HeLa cells that Hsp90 inhibition can prevent the formation of a proposed hetero-complex involving IKK and Cdc37/Hsp90, reducing the size of the complex and preventing activation of NF- κ B²⁶. It has also been observed in alveolar epithelial cells that treatment with Hsp90 inhibition induces dissociation of Hsp90 from the IKK complex, rendering the complex detergent insoluble and the NF- κ B system impassive to cytokine stimulation³⁷.

It therefore seems likely that Hsp90 inhibitors repress TLR4-mediated NF- κ B activity primarily through IKK. The breakdown of all the major subunits of IKK by AT13387 and 17-DMAG following hyaluronan stimulation in this study would support this hypothesis. Although AT13387 was significantly more effective at reducing hyaluronan-mediated NF- κ B activity, it does not appear to act through different mechanisms from 17-DMAG. However, as it is a smaller molecule Hsp90 inhibitor it may be more efficacious in this respect.

References

- Lameire, N. H. *et al.* Acute kidney injury: an increasing global concern. *Lancet* **382**, 170–179 (2013).
- Sharfuddin, A. A. & Molitoris, B. A. Pathophysiology of ischemic acute kidney injury. *Nat Rev Nephrol* **7**, 189–200 (2011).
- Murugan, R. & Kellum, J. A. Acute kidney injury: what's the prognosis? *Nat Rev Nephrol* **7**, 209–217 (2011).
- Harrison, E. M. *et al.* Heat shock protein 90-binding agents protect renal cells from oxidative stress and reduce kidney ischemia-reperfusion injury. *Am J Physiol Renal Physiol* **295**, F397–405 (2008).
- Wu, H. *et al.* TLR4 activation mediates kidney ischemia/reperfusion injury. *J Clin Invest* **117**, 2847–2859 (2007).
- Hayden, M. S. & Ghosh, S. Signaling to NF-kappaB. *Genes Dev* **18**, 2195–2224 (2004).
- Salminen, A., Paimela, T., Suuronen, T. & Kaarniranta, K. Innate immunity meets with cellular stress at the IKK complex: regulation of the IKK complex by HSP70 and HSP90. *Immunol Lett* **117**, 9–15 (2008).
- Mahadevan, D. *et al.* Activity of AT13387, a novel, non-ansamycin inhibitor of heat shock protein 90, against gastrointestinal stromal tumors (GIST). *Journal of Clinical Oncology* **31**, abstr 105 (2013).
- Woodhead, A. J. *et al.* Discovery of (2,4-dihydroxy-5-isopropylphenyl)-[5-(4-methylpiperazin-1-ylmethyl)-1,3-dihydroisindol-2-yl]methanone (AT13387), a novel inhibitor of the molecular chaperone Hsp90 by fragment based drug design. *J Med Chem* **53**, 5956–5969 (2010).
- Hesketh, E. E. *et al.* Renal ischaemia reperfusion injury: a mouse model of injury and regeneration. *J Vis Exp* (2014).
- Borner, U., Szasz, G., Bablok, W. & Busch, E. W. [A specific fully enzymatic method for creatinine: reference values in serum (author's transl)]. *J Clin Chem Clin Biochem* **17**, 679–682 (1979).
- Schneider, C. A., Rasband, W. S. & Eliceiri, K. W. NIH Image to ImageJ: 25 years of image analysis. *Nat Methods* **9**, 671–675 (2012).
- Faul, F., Erdfelder, E., Lang, A. G. & Buchner, A. G*Power 3: a flexible statistical power analysis program for the social, behavioral, and biomedical sciences. *Behav Res Methods* **39**, 175–191 (2007).
- Tian, B., Nowak, D. E., Jamaluddin, M., Wang, S. & Brasier, A. R. Identification of direct genomic targets downstream of the nuclear factor-kappaB transcription factor mediating tumor necrosis factor signaling. *J Biol Chem* **280**, 17435–17448 (2005).
- Zhou, A., Scoggin, S., Gaynor, R. B. & Williams, N. S. Identification of NF-kappa B-regulated genes induced by TNFalpha utilizing expression profiling and RNA interference. *Oncogene* **22**, 2054–2064 (2003).
- Miura, M., Fu, X., Zhang, Q. W., Remick, D. G. & Fairchild, R. L. Neutralization of Gro alpha and macrophage inflammatory protein-2 attenuates renal ischemia/reperfusion injury. *Am J Pathol* **159**, 2137–2145 (2001).
- Linkermann, A. *et al.* Rip1 (receptor-interacting protein kinase 1) mediates necroptosis and contributes to renal ischemia/reperfusion injury. *Kidney Int* **81**, 751–761 (2012).
- Hughes, M. A. *et al.* MyD88-dependent signaling contributes to protection following Bacillus anthracis spore challenge of mice: implications for Toll-like receptor signaling. *Infect Immun* **73**, 7535–7540 (2005).
- Thomas, P. & Smart, T. G. HEK293 cell line: a vehicle for the expression of recombinant proteins. *Journal of pharmacological and toxicological methods* **51**, 187–200 (2005).
- Meldrum, K. K. *et al.* Profibrotic effect of interleukin-18 in HK-2 cells is dependent on stimulation of the Toll-like receptor 4 (TLR4) promoter and increased TLR4 expression. *J Biol Chem* **287**, 40391–40399 (2012).
- Israel, A. The IKK complex, a central regulator of NF-kappaB activation. *Cold Spring Harb Perspect Biol* **2**, a000158 (2010).
- Dello Russo, C. *et al.* The heat-shock protein 90 inhibitor 17-allylamino-17-demethoxygeldanamycin suppresses glial inflammatory responses and ameliorates experimental autoimmune encephalomyelitis. *J Neurochem* **99**, 1351–1362 (2006).
- Poulaki, V. *et al.* Inhibition of Hsp90 attenuates inflammation in endotoxin-induced uveitis. *FASEB J* **21**, 2113–2123 (2007).
- Lewis, J. *et al.* Disruption of hsp90 function results in degradation of the death domain kinase, receptor-interacting protein (RIP), and blockage of tumor necrosis factor-induced nuclear factor-kappaB activation. *J Biol Chem* **275**, 10519–10526 (2000).
- Malhotra, V., Shanley, T. P., Pittet, J. F., Welch, W. J. & Wong, H. R. Geldanamycin inhibits NF-kappaB activation and interleukin-8 gene expression in cultured human respiratory epithelium. *Am J Respir Cell Mol Biol* **25**, 92–97 (2001).
- Chen, G., Cao, P. & Goeddel, D. V. TNF-induced recruitment and activation of the IKK complex require Cdc37 and Hsp90. *Mol Cell* **9**, 401–410 (2002).
- Broemer, M., Krappmann, D. & Scheidereit, C. Requirement of Hsp90 activity for IkappaB kinase (IKK) biosynthesis and for constitutive and inducible IKK and NF-kappaB activation. *Oncogene* **23**, 5378–5386 (2004).

28. Lee, K. H., Jang, Y. & Chung, J. H. Heat shock protein 90 regulates IkappaB kinase complex and NF-kappaB activation in angiotensin II-induced cardiac cell hypertrophy. *Exp Mol Med* **42**, 703–711 (2010).
29. Chen, J. *et al.* Early interleukin 6 production by leukocytes during ischemic acute kidney injury is regulated by TLR4. *Kidney Int* **80**, 504–515 (2011).
30. Rusai, K. *et al.* Toll-like receptors 2 and 4 in renal ischemia/reperfusion injury. *Pediatr Nephrol* **25**, 853–860 (2010).
31. Pulsikens, W. P. *et al.* Toll-like receptor-4 coordinates the innate immune response of the kidney to renal ischemia/reperfusion injury. *PLoS One* **3**, e3596 (2008).
32. Liu, M. *et al.* Protective effects of Toll-like receptor 4 inhibitor eritoran on renal ischemia-reperfusion injury. *Transplant Proc* **42**, 1539–1544 (2010).
33. Feng, B. *et al.* Small interfering RNA targeting RelB protects against renal ischemia-reperfusion injury. *Transplantation* **87**, 1283–1289 (2009).
34. Cao, C. C. *et al.* *In vivo* transfection of NF-kappaB decoy oligodeoxynucleotides attenuate renal ischemia/reperfusion injury in rats. *Kidney Int* **65**, 834–845 (2004).
35. Wan, X. *et al.* Small interfering RNA targeting IKKbeta prevents renal ischemia-reperfusion injury in rats. *Am J Physiol Renal Physiol* **300**, F857–863 (2011).
36. Chen, L. W. *et al.* The two faces of IKK and NF-kappaB inhibition: prevention of systemic inflammation but increased local injury following intestinal ischemia-reperfusion. *Nat Med* **9**, 575–581 (2003).
37. Pittet, J. F. *et al.* Stress-induced inhibition of the NF-kappaB signaling pathway results from the insolubilization of the IkappaB kinase complex following its dissociation from heat shock protein 90. *J Immunol* **174**, 384–394 (2005).

Acknowledgements

This study was funded by grants from Tenovus Scotland and The Royal College of Surgeons Edinburgh. The Maurice Wohl Research Fellowship from the Royal College of Surgeons Edinburgh, The Mason Medical Research Trust Fellowship and a Medical Research Council Clinical Research Training Fellowship has supported S.O.N.

Author Contributions

S.O.N. designed the experiments, performed the experiments, analysed data and drafted the manuscript. D.H. and G.T. contributed to the experimental work, drafting of the manuscript and revised the manuscript critically for important intellectual content. L.P.M. and K.D. supervised the experimental work of D.H. and G.T., contributed to the drafting of the manuscript and revised the manuscript critically for important intellectual content. J.H., J.A.R., S.J.W. and E.M.H. supervised the experimental work of S.O.N. and contributed substantially to conception of the study, design of the experiments, analysis of data, drafting of the manuscript and revision of the manuscript critically for important intellectual content. Final approval of the version to be published was obtained. All authors agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: O'Neill, S. *et al.* Heat shock protein 90 inhibition abrogates TLR4-mediated NF- κ B activity and reduces renal ischemia-reperfusion injury. *Sci. Rep.* **5**, 12958; doi: 10.1038/srep12958 (2015).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>